

Dissecting the role of *WBPI* in preimplantation embryonic development: Insights into hippo signaling and trophectoderm formation in the bovine

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A SNP in WW binding domain protein (*WBPI*) has been linked to cow conception rate, a fertility trait in dairy cattle, and embryonic development in vitro. *WBPI* interacts with the WW domains and serves as a mediator of protein-protein interactions. In the embryo, the Hippo signaling pathway is composed of many proteins that contain WW domains, including the Yes-associated protein (YAP) which has been associated with trophectoderm (TE) formation. Previous research from our group has shown that when the expression of *WBPI* is reduced, blastocyst rate decreases and so does its number of trophectoderm cells. This suggests that *WBPI* may be involved in trophectoderm differentiation, however, the exact mechanism of action of *WBPI* in the embryo is still unknown. The objective of this study was to determine how the functional ablation of *WBPI* affects preimplantation embryonic development. To test this, a CRISPR-CAS9 system was used to ablate *WBPI*. Briefly, two guides spanning intron 1 and exon 2 were annealed to a tracer and to CAS9mRNA to form a solution of 62.5 ng/ul of each guide and 125 ng/ul of CAS9mRNA. The complex of guideRNA-CAS9 was electroporated into presumptive zygotes 10 hours post-insemination for treated embryos and just Cas9mRNA for controls. Zygotes were cultured in SOF-BEII media at 38.5°C with 5%CO₂ and 5%O₂. Cleavage was measured at day 3 and blastocyst at day 7.5 of development. To assess trophectoderm formation, immunolocalization of CDX2, phosphorylated YAP, and YAP translocation to the nucleus was measured at the 8-16 cell and morula stages. Images were analyzed using ImageJ v1.44. Cleavage was not different ($P = 0.1282$) between *WBPI*-null and control embryos (LSMEANS \pm SEM: 74.84 \pm 3.27% and 67.44 \pm 3.64%, respectively). Control embryos had a blastocyst rate of 31.60 \pm 3.4% (LSMEANS \pm SEM), however, *WBPI*-null embryos did not produce blastocyst, and embryos were arrested at the morula stage ($P < 0.0001$). At the 8-16 cell stage, there were fewer ($P = 0.0028$) nuclear YAP-positive cells on *WBPI*-null embryos (53.13 \pm 6.32% of total cells), compared to controls (82.65 \pm 3.83% of total cells). In addition, the mean intensity of phosphorylated YAP was higher ($P = 0.0246$) in *WBPI*-null embryos (40.06 \pm 3.21) compared to controls (24.76 \pm 2.78). There was no difference ($P = 0.866$) in the number of CDX2-positive cells between groups at this stage. In contrast, at the morula stage, *WBPI*-null embryos had fewer ($P = 0.017$) cells with nuclear YAP (56 \pm 3.75%), compared to control embryos (81.87 \pm 2.95%). In addition, *WBPI*-null morulas had a reduced ($P = 0.0195$) number of CDX2-positive cells (49.14% \pm 3.78%) than control (64.33 \pm 3.66%). These results indicate that *WBPI* has an important role in the hippo signaling pathway by regulating YAP. The absence of *WBPI* causes increased YAP phosphorylation and degradation in the cytoplasm impairing TE differentiation which results in embryonic arrest before the blastocyst stage. Further research is required to elucidate if *WBPI* acts in other proteins from the hippo signaling or how it regulates YAP function. Funding provided by USDA-AFRI 2022-6701538938.