## Hypoxia-Related Regulation of Luteal Function: New Insights from Gene Expression and LPS Challenge Studies in Cattle

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Considering the role of the corpus luteum (CL) in the reproductive performance of cattle, comprehending its formation, maintenance and regression remains of utmost clinical importance. Reduced oxygen content has emerged as a significant factor influencing these processes. In this study, we explored the dynamics of CL functionality, investigating gene expression patterns of hypoxia-mediating and regulating factors during the luteal phase, and in response to Escherichia coli lipopolysaccharide (LPS) challenge, as well as inflammation related factors. LPS, a gram-negative bacterial endotoxin, frequently causes infertility and subfertility in livestock. When administered at 0.5µg/kg i.v., LPS affects CL function, decreases its steroidogenic capacities, and triggers systemic responses, without causing a lifethreatening risk. Here: (1) CL samples were collected during different luteal stages (early, mid, and late); (2) CL biopsies from synchronized cows were obtained on day 10 of the first estrous cycle 12h after being treated i.v. with LPS or saline, and on day 10 of the subsequent cycle. Semi-quantitative real-time TaqMan PCR was performed to assess gene expression. Nonradioactive in situ hybridization (ISH) localized hypoxia inducible factor (HIF) 1a and its regulators. The transcript levels of *HIF1A* and its oxygen-sensitive regulators: PHD1 (*EGLN2*), PHD2 (EGLN1), PHD3 (EGLN3), FIH (HIF1AN), and VHL were assessed. The results demonstrated dynamic changes in gene expression patterns during the luteal lifespan, indicating active regulation of HIF1 $\alpha$  transcriptional and functional availability. The luteal development was accompanied by increasing HIF1A transcription while it decreased in the late luteal phase. Its elevated levels at mid-luteal stage were accompanied by time-dependent expression of EGLN2, HIF1AN and VHL. In regressing CL, VHL and EGLN3 were significantly downregulated. While the LPS challenge affected only the expression of EGLN2 and EGLN3 in opposite directions, with EGLN2 being downregulated and EGLN3 upregulated, we observed a significant increase in the response of endothelial cell-specific pro-inflammatory factors ICAM1 (ICAM1) and NFxB (NFKB2), suggesting that LPS-induced vascular inflammation occurs, with concomitant hypoxia-related effects, possibly modulated through EGLN2 and -3 expression. ISH revealed compartmentalization of signals regarding the distribution of HIF1 $\alpha$  and its regulators within the bovine CL, emphasizing the cell-specific regulation of hypoxia-related factors. Large luteal cells presented predominant signals for HIF1A. These cells were shown to exhibit higher basal progesterone (P4) production, than small luteal cells and less responsiveness to LH stimulation, implying that the basal production of P4 in the CL and the constant provision of P4 remain under the control of HIF1 $\alpha$  in large luteal cells. HIF1A also showed signals in capillary vessels, mostly in endothelial cells, as well as in the media of smaller vessels, where it was colocalized with the other regulatory factors. EGLN2, prevailed in vessels, whereas in luteal cells it was mostly targeted to large cells, thereby colocalized with HIF1A. EGLN1 was abundantly present in larger vessels and ubiquitously distributed in small and large luteal cells, although it appeared weaker than in vessels. EGLN3 and VHL followed the localization pattern of EGLN1, but appeared to prevail in the vessels. In conclusion, this study offers new insights into hypoxia-dependent regulation of luteal function, including the spatiotemporal distribution of signals encoding for HIF1 $\alpha$ regulatory factors. It marks the first demonstration of HIF1 $\alpha$  expression in the context of its regulators and reveals their dynamic cellular compartmentalization. Additionally, it broadens our understanding of the LPS-mediated inflammatory responses in the CL, implicating possible hypoxia-mediated effects.